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PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA, vol. 79, no. 9, May 1982, Baltimore J. LIEMAN-HURWITZ et al. "Human cytoplasmic superoxide dismutase cDNA clone: a probe for studying the molecular biology of Down syndrome" pages 2808-2811

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DescriptionField of the Invention

5 Superoxide dismutase ("SOD") is in fact a variety of different enzymes found in most living organisms. One function in mammals is to destroy superoxide, a material naturally produced during phagocytosis. The superoxide dismutases are characterized in families based on the metal associated with the enzyme, where the metals vary amongst iron, manganese, copper and copper-zinc. Superoxide dismutase, e.g., from bovine liver, has found clinical use, particularly as an anti-inflammatory agent in mammals including
 10 humans. Other utilities include scavenging superoxide anions due to exposure of a host to various superoxide-inducing agents, e.g. radiation, paraquat, etc.; prophylaxis or therapy for certain degenerative diseases, e.g., emphysema; food preservation; and the like.

It is therefore important that stable supplies of physiologically acceptable superoxide dismutase be made available, particularly for use in vivo as an anti-inflammatory agent or for other therapeutic purposes.
 15 For human application it would be preferable to employ the homologous enzyme to prevent or minimize possible immune response. By employing recombinant DNA techniques, there is the opportunity to produce products efficiently, which have the desired biological activities of superoxide dismutase, such as immunological and enzymatic activities.

20 Description of the Prior Art

The amino acid sequence of human erythrocyte Cu-Zn superoxide dismutase is described in Jabusch et al., Biochemistry (1980) 19:2310-2316 and Barra et al., FEBS Letters (1980) 120:53-55. Bovine erythrocyte Cu-Zn SOD is described by Steinman et al., J. Biol. Chem. (1974) 249:7326-7338. A SOD-1 cDNA
 25 clone is described by Lieman-Hurwitz et al., Proc. Natl. Acad. Sci. USA (1982) 79:2808-2811. Concerning the effect on efficiency of translation of varying the untranslated region upstream from the initiation codon, see Gheysen et al., Gene (1982) 17:55-63; Thunmel et al., J. Virol. (1981) 37:683-697; and Matteucci and Heyneker, Nucl. Acids Res. (1983) 11:3113-3121.

30 SUMMARY OF THE INVENTION

Polypeptides acetylated at their N-terminus and methods for producing such acetylated polypeptides are provided. By providing a particular acetylation signal sequence at the 5'-end of the structural gene for a desired polypeptide, the N-terminal amino acid will be acetylated when the gene is expressed in yeast. The
 35 acetylation signal sequence encodes for at least the first two N-terminal amino acids, where the first amino acid is either alanine or glycine, and the second amino acid is a polar amino acid, usually being threonine, serine or aspartate. Acetylation of human superoxide dismutase produced in yeast is demonstrated when the first two amino acids are alanine and threonine, respectively.

40 BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 indicates the DNA linker sequence and a flow diagram showing its use;
 Figs. 2 and 3 are flow diagrams indicating the preparation of plot5/SOD.
 Fig. 4 indicates the sequence of both the coding strand of human SOD cDNA (5'→3') and the resultant
 45 translation product.
 Fig. 5 illustrates the sequence of the isolated human SOD gene described in the Experimental section hereinafter.
 Fig. 6 is a restriction map of the isolated human SOD gene described in the Experimental section hereinafter.

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DESCRIPTION OF SPECIFIC EMBODIMENTS

Methods and compositions are provided for the efficient expression of polypeptides demonstrating the biological activities of human Cu-Zn superoxide dismutase ("hSOD"). The methods employ a DNA
 55 sequence ("hSOD gene") encoding a substantial portion of the amino acid sequence of hSOD in conjunction with a translational initiation region optimized for expression in the expression host. The hSOD gene is inserted into an appropriate vector for expression in a host, conveniently under conditions which allow for secretion, so as to harvest the SOD product from the extracellular medium.

Methods and compositions are also provided for the N-terminal acetylation of hSOD and other polypeptides. Hereinafter, acetylation refers to addition at the amino terminus of polypeptides and proteins in contrast to modification of amino acid side chains, e.g., lysine, as is also observed naturally. Acetylation of polypeptides and proteins is useful for a number of reasons. Where the natural condition of the polypeptide includes acetylation, as is the case for cytoplasmic hSOD, methods of expression which include acetylation provide a product having the desired natural structure and conformation. Where the product finds pharmaceutical and/or in vitro or in vivo diagnostic use, the acetylated material will minimize or eliminate immunogenicity when administered to a host and/or exposed to biological samples. Also, acetylated polypeptides are likely to be more stable and resistant to degradation by proteases and thus enjoy a prolonged existence in the cell, blood or body and tissue fluids.

The structural gene for hSOD or other polypeptide includes an acetylation signal sequence at the 5'-end thereof, which signal sequence causes a yeast expression host to effect acetylation. The acetylation signal sequence encodes at least the first two N-terminal amino acids in the polypeptide. The first amino acid will be either alanine, glycine or serine, while the second amino acid will be a polar or aromatic amino acid, usually being threonine, serine, aspartate or phenylalanine.

The amino acids may be the natural N-terminal amino acids normally present in the polypeptide to be expressed. This is the case with hSOD where the first two amino acids are alanine and threonine, respectively. Other naturally-acetylated proteins which may be expressed and acetylated in yeast include:

20

<u>Protein</u>	<u>Source</u>	<u>Signal Sequence</u>
Cytochrome C	Human, Rhesus Monkey, Dog, Horse, etc.	GLY-ASP
Cytochrome C	Castor, Sesame, Mung-bean, etc.	ALA-SER
Glutamate dehydrogenase	Neurospora	SER-ASN
Calmodulin	Pig	SER-ALA
Myosin (light chain A2)	-----	SER-PHE
ADH	Drosophila	SER-PHE

40

The present invention is also useful for acetylating polypeptides and proteins which are not naturally acetylated. Acetylation may be achieved by joining the acetylation signal sequence to the 5'-end of the structural gene for the polypeptide. The acetylation signal sequence will encode for at least two amino acids (as described above), and may encode up to ten or more amino acids, preferably fewer than five amino acids. Fewer added amino acids is usually desirable to limit interference with or loss of a desired activity of the polypeptide. Conveniently, the signal sequence may be synthesized and joined to the structural gene using well known techniques.

As an alternative to adding the acetylation signal sequence to the structural gene, it will sometimes be possible to modify the 5'-end of the structural gene to substitute one or both of the first two amino acids of the polypeptide. Such modification may be accomplished by a variety of conventional methods. For example, the structural gene may be restricted near its 5'-end to remove a known number of nucleotides. A synthetic oligonucleotide may then be joined to the cohesive end remaining after restriction. The oligonucleotide will restore and substitute the base pairs as necessary to provide the desired acetylation signal sequence. Alternatively, site-specific mutagenesis employing, e.g., phage M13, can be used to effect an appropriate modification to the 5'-end of the structural gene.

In order to prepare hSOD, it is necessary to have a DNA sequence which encodes for hSOD. One manner of achieving such sequence, is to clone cDNA from messenger RNA from cells which produce hSOD. Conveniently, human liver cells may be used for this purpose. After the cDNA is cloned, where the

DNA coding sequence is unknown, but at least a partial amino acid sequence is known, one may then screen the cDNA with mixtures of probes having all of the possible variations of nucleotides encoding for the particular series of amino acid residues. The choice of the residues for which the sequence encodes is somewhat arbitrary, although the residues chosen will usually be selected to minimize the number of
 5 different sequences which must be synthesized.

For hSOD, conveniently a DNA sequence encoding for at least the amino acid residues 19 to 24 can be used, particularly a probe having at least about 15 bases and not more than about 20 bases, more conveniently about 17 bases. One may then restriction enzyme digest the clones which appear to hybridize with the labeled probes, fractionate the DNA fragments and repeat the hybridization, particularly by
 10 employing a second series of probes which hybridize to DNA sequences encoding for a different series of amino acid residues in hSOD. Conveniently, these amino acid residues may be 109 to 114. One or more clones may be found which are positive to both probes and these may be used as a source for cDNA encoding for at least a substantial proportion of hSOD.

Quite surprisingly, it was found that the amino acid sequences which have been published for hSOD
 15 differed in a significant number of residues from the amino acid sequence encoded for by the cDNA. Specifically, where the two published sequences differed (Jabusch et al., *Biochemistry* (1980) 19:2310-2316 and Barra et al., *FEBS Letters* (1980) 120:153-156), the correct assignments are: residue 11, aspartate; residue 17, isoleucine; residue 26, asparagine; residue 49, glutamate; residue 52, aspartate; residue 53, asparagine; residue 92, aspartate; residue 98, serine (see Fig. 4).

Because of the uncertainties of the effect on translation of the separation between the ribosomal binding site and the translational initiation codon, normally AUG, the subject method provides a technique for varying the distance and nucleotides separating the ribosomal binding site from the initiation codon. Usually, there are from about 6 to 15, more usually about 6 to 12 nucleotides in the spacer between the ribosomal binding site and initiation codon. As the base sequence downstream from the initiation site may also affect
 25 translation efficiency, the subject method also provides for variation of nucleotide sequence (but not length) within the initial several 5'-codons of the polypeptide as permitted by the redundancy constraints of the genetic code. Such degeneracy may extend up to 4 codons, more usually 2 codons, downstream from the initiation site.

A plurality of linkers are prepared where at least 2 nucleotides, usually at least 3 nucleotides, and not
 30 more than 10 nucleotides, usually not more than about 6 nucleotides, are varied to include members having each of the 4 nucleotides if within the spacer or 2, 3, or 4 nucleotides as permitted by genetic code redundancy if within the structural gene for the polypeptide. In addition, the linkers are prepared, having differing numbers of nucleotides, so as to provide a group of linkers differing not only in the sequence, but also in length. The difference in length can be achieved by removal of portions of the support during the
 35 linker synthesis and, if appropriate, continuing synthesis at a subsequent stage, so as to provide for linkers having a graduated number of sequence lengths. Usually, the mixture of linkers will vary in length by at least one nucleotide and not more than over a range of six nucleotides, usually not more than four nucleotides.

This can be conveniently illustrated where the absent bases are at the terminus. After each stage, a
 40 portion of the support is removed and the synthesis continued with the strands bound to the support, providing all four nucleotides (dNTP) at each stage. These single strands will then be hybridized to a single strand which is complementary in part, where the variable region will be an overhang. Thus, one will achieve a graduated series of linkers having overhangs differing in both their nucleotide sequences and lengths. At an appropriate point during subsequent hybridization, ligation or cloning operations the overhang
 45 region(s) is filled in to provide double-stranded material amenable to further manipulation. This is usually and preferably performed in vitro, e.g., using the Klenow fragment of DNA polymerase I; alternatively, in certain constructs the overhang could be cloned as a single strand with filling in occurring in vivo in the transformed or transfected host. Hybridization to a complementary strand can be achieved by having a 5'-sequence upstream from the variable nucleotide series which is complementary to a sequence present in
 50 the terminal sequence to which the linker is to be joined. The missing bases may then be filled in vitro or in vivo.

The linkers include within their sequence, at least a portion of the region between the ribosomal binding site and the initiation codon, preferably the nucleotides proximal to the initiation codon. The linker may also include the initiation codon and portions of the structural gene, the ribosomal binding site, and bases
 55 upstream from the ribosomal binding site, which may or may not include transcriptional regulatory sequences.

Usually the linker will be at least about 5 bases, more usually at least about 20 bases, and usually not exceeding about 200 bases, more usually not exceeding about 100 bases. Where the linker is greater than

about 35 bases, it will usually be assembled by employing single stranded sequences of from about 10 to 35 bases, which have homology with only a part of a complementary strand, thus providing for complementary overlapping sequences with overhangs, so that the various single strands can be hybridized, ligated and the degenerate and/or variable length overhang filled in as indicated above to produce the desired
 5 linker having cohesive and/or blunt ends.

Where the structural gene has a convenient restriction site, usually not more than about 50 bases downstream from the initiation codon, a fragment containing the structural gene may be restricted and joined to a complementary cohesive terminus of the linker or may be filled in to provide a blunt-end terminus, which blunt end may be ligated to a blunt end of the linker. The linker is devised to ensure that
 10 the structural gene is complete and in reading frame with the initiation codon.

As indicated, in preparing the linker, one provides that there are a series of linkers which have a randomized series of nucleotides, that is, each of the four possible nucleotides in the coding strand (subject to the provision of genetic code limitations indicated above) and which are graduated in size, lacking one or more of the nucleotides defining the region intermediate or bridging the ribosomal binding site and initiation
 15 codon. These linkers which are prepared from single strands may be joined to other single or double DNA strands to provide for extended linkers, which may include not only the ribosomal binding site, but bases upstream from the ribosomal binding site. Alternatively, the linkers may be relatively small, beginning at a site internal to or adjacent to the ribosomal binding site and extending downstream to a site at the initiation codon or internal to the structural gene.

While the particular order of joining the various fragments to produce the constructs of this invention will usually not be critical, conveniently, the structural gene may be first joined to the linker. This DNA construct will include not only the structural gene, but also the ribosomal binding site and any additional nucleotides upstream from the ribosomal binding site. In addition, there will be substantial variety in the nucleotides and numbers of nucleotides between the ribosomal binding site and initiation codon. The subject DNA construct
 25 is inserted into an appropriate expression vector which has the necessary transcriptional initiation regulatory sequences upstream, as well as transcriptional termination regulatory sequences downstream from the insertion site of the subject DNA construct. Thus, the linker will be flanked at the 5'-end with transcriptional initiation regulatory signal sequences and at the 3'-end with at least a portion of a coding region and transcriptional and translational termination sequences. (5'- and 3'-intend the direction of transcription.)

After preparing the plasmid or viral DNA for introduction into an appropriate host (usually including at an appropriate stage in the manipulations filling in of the variable overhang region), the host is transformed or transfected, respectively, cloned, the clones streaked and individual clones selected for efficient expression by assaying for production of the desired product, e.g., hSOD. The number of clones to be screened to determine the various levels of production of the product will depend upon and be proportional to the
 35 degree of length variability and sequence degeneracy introduced into the synthetic linker. As exemplified in the present embodiment, with 4 length variables and 4-fold sequence degeneracy at each of 6 nucleotides in the linker, the number of possible recombinant sequences is 5440. Usually at least a few hundred, preferably several thousand or more, clones will be screened. Screening can be efficiently performed using Western blots (antibody detection of product) of host cell colonies or viral plaques
 40 transferred to filters of nitrocellulose or other suitable material. Alternatively, using electrophoresis and providing for a plurality of lanes, where each lane is an individual clone, an immediate and direct comparison can be made of which clones are most efficient in expression by visualization of staining intensity, autoradiography or Western blotting of the product band. This screen will usually be sufficient, although more quantitative immunoassays or enzyme assays can be employed, as appropriate.

If desired, the construct can be transferred to a different host which recognizes the regulatory signals of the expression construct or the expression construct modified by introduction at appropriate sites of necessary regulatory signals to provide for efficient expression in an alternative host.

If desired, the hSOD gene may be joined to secretory leader and processing signals to provide for secretion and processing of the hSOD. Various secretory leader and processing signals have been
 50 described in the literature. See for example, U.S. Patent Nos. 4,336,336 and 4,338,397, as well as copending application Serial Nos. 522,909, filed August 12, 1983 and 488,857, filed April 26, 1983.

Of particular interest as hosts are unicellular microorganism hosts, both prokaryotes and eukaryotes, such as bacteria, algae, fungi, etc. In particular, E. coli, B. subtilis, S. cerevisiae, Streptomyces, Neurospora may afford hosts.

A wide variety of vectors are available for use in unicellular microorganisms, the vectors being derived
 55 from plasmids and viruses. The vectors may be single copy or low or high multicopy vectors. Vectors may serve for cloning and/or expression. In view of the ample literature concerning vectors, commercial availability of many vectors, and even manuals describing vectors and their restriction maps and char-

acteristics, no extensive discussion is required here. As is well-known, the vectors normally involve markers allowing for selection, which markers may provide for cytotoxic agent resistance, prototrophy or immunity. Frequently, a plurality of markers are present, which provide for different characteristics.

In addition to the markers, vectors will have a replication system and in the case of expression vectors, will usually include both the initiation and termination transcriptional regulatory signals, such as promoters, which may be single or multiple tandem promoters, an mRNA capping sequence, a TATA box, enhancers, terminator, polyadenylation sequence, and one or more stop codons associated with the terminator. For translation, there will frequently be a ribosomal binding site as well as one or more stop codons, although usually stop codons will be associated with a structural gene. Alternatively, these regulatory sequences may be present on a fragment containing the structural gene, which is inserted into the vector.

Usually, there will be one or more restriction sites conveniently located for insertion of the structural gene into the expression vector. Once inserted, the expression vector containing the structural gene may be introduced into an appropriate host and the host cloned providing for efficient expression of hSOD.

In some instances, specialized properties may be provided for the vector, such as temperature sensitivity of expression, operators or activators for regulation of transcription, and the like. Of particular interest is the ability to control transcription by exogenous means, such as temperature, inducers, corepressors, etc., where transcription can be induced or repressed by an exogenous compound, usually organic.

Where the hSOD is made intracellularly, when the cell culture has reached a high density, the cells may be isolated, conveniently by centrifugation, lysed and the hSOD isolated by various techniques, such as extraction, affinity chromatography, electrophoresis, dialysis, or combinations thereof. Where the product is secreted, similar techniques may be employed with the nutrient medium, but the desired product will be a substantially higher proportion of total protein in the nutrient medium than in the cell lysate.

The hSOD which is formed has substantially the same amino acid sequence as the naturally occurring human superoxide dismutase, usually differing by fewer than 5 amino acids, more usually differing by fewer than 2 amino acids. The recombinant hSOD ("r-hSOD") displays substantially the same biological properties as naturally occurring hSOD. The biological properties include immunological properties, where antibodies raised to authentic hSOD cross-react with r-hSOD. Furthermore, in common bioassays employed for hSOD, the r-hSOD product demonstrates a substantial proportion, usually at least about 10%, preferably at least about 50%, more preferably at least about 80%, of the enzymatic activity of the authentic hSOD, based on weight of protein. An illustrative assay technique is described by Marklund and Marklund, Eur. J. Biochem. (1974) 47:469-474.

The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

Molecular Cloning of hSOD cDNA

Total RNA was prepared from an adult human liver by the guanidinium thiocyanate/lithium chloride method (Cathala et al., DNA (1983) 2:329-335). polyA RNA was used to synthesize double-stranded cDNA (Maniatis et al., Molecular Cloning, 213-242, Cold Spring Harbor, 1982) and this was passed over a Sepharose CL4B column to enrich for cDNAs of greater than 350bp (Fiddes and Goodman, Nature (1979) 281:351-356). The cDNA was inserted at the PstI site of plot4, a pBR322 derivative having the following sequence replacing the PstI-EcoRI site.

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1  PstI      HinfI                AluI
   GGTGAATCCGTAATCATGGTCATAGCTGTTTCCTGTGTGAAATTGTTATCCGCTC
   ACGTCCACTTAGGCATTAGTACCAGTATCGACAAAGGACACACTTTAACAATAGCCGAG
      HphI

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60  HindIIIAluI
   ACAATTCCACACATTATACGAGCCGATGATTAATTGTCAACAGCTCATTTGAGAATATTT
   TGTTAAGGTGTGTAATATGCTCGGCTACTAATTAACAGTTGTCGAGTAAAGTCTTATAAA

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120 EcoRI
   GCCAGAACCGTTATGATGCGG
   CCGTCTTGGCAATACTACGCCCTTAA

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The cDNA insertion employed the oligo-dG:dC tailing method (Maniatis et al., *supra*). *E. coli* strain D1210 was transformed with this mixture and transformants selected on L-agar containing 10 µg/ml tetracycline (Kushner, S.R. (1978) In: *Genetic Engineering*, eds. Boyer, H.B. and Nicosia, S., (Elsevier/North Holland, Amsterdam) p. 17). Plasmid DNA constituting a liver cDNA library was prepared (Maniatis et al., *Molecular Cloning*, pp. 88-94, Cold Spring Harbor 1982) directly from approximately 62,000 recombinant colonies plated at a density of approximately 3,000 colonies per 9cm diameter Petri dish.

Isolation of r-hSOD Clones

Strain D1210 was retransformed with the liver cDNA library and about 40,000 clones were grown on nine 14cm diameter Petri dishes. After transfer of the colonies to nitrocellulose paper and chloramphenicol amplification of plasmid DNA, the cells were lysed and the filters prepared for hybridization (Ish-Horowitz and Burke, *Nucleic Acids Research* (1981) 9:2989-2998). Oligonucleotide probes were employed for screening by hybridization, with the probes consisting of enzymatically-radiolabeled, chemically-synthesized DNA molecules complementary to the mRNA encoding amino acid residues 19 to 24 of the protein (Jabusch et al., *supra*; Barra et al., *supra*); the mixture had the following sequences:

3' TTA AAA CTT GTT TTT CT 5'
G G C C C

where all of the indicated possibilities for encoding the peptide sequence were prepared (32-fold degenerate).

The probes were labeled with ³²P to a specific activity of 1-3x10⁸ cpm/µg and Millipore (0.45µm) filtered before use. Filters were prehybridized for 6hrs at 30 °C in 4x SSC, 2x Denhardt's solution, 40mM sodium phosphate, pH 7.5, 300µg/ml sonicated salmon testes DNA. Hybridization was for 20hrs at 30 °C in the same solution containing 2x10⁶ cpm/ml hSOD DNA probe (residues 19-24). Filters were washed in 4x SSC, once for 15min at r.t. and twice for 15min at 30 °C, blotted dry and autoradiographed with an intensifying screen for 24hrs at -70 °C.

Areas on the master plates that corresponded to duplicate positive signals were picked into L-broth and plasmid DNA prepared by the miniscreen procedure (Maniatis et al., *Molecular Cloning*, 178, 368-369, Cold Spring Harbor 1982). This DNA was cut with PstI and subjected to Southern blot analysis (Southern, J. Mol. Biol. (1975) 98:503-517) hybridizing initially with the previous labeled probes (amino acid residues 19-24) and then with additional radiolabeled probes derived from amino acid residues 109-114 and having the following sequences (all possible variations, 72-fold degenerate) present as a mixture:

3' CTA GTA ACA TAA TAA CC 5'
G G G G G
T T

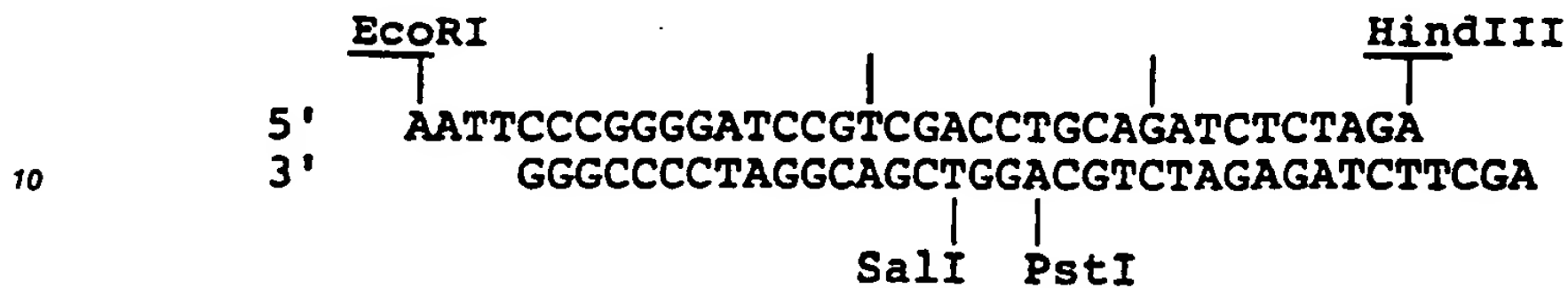
One plasmid pool (pSOD1) contained a cDNA insert of 520bp that hybridized with both probes and after colony purification, plasmid DNA was prepared from this clone and sequenced by the method of Maxam and Gilbert (*Proc. Natl. Acad. Sci. USA* (1977) 74:560-564) with the results shown in Fig. 4. The hSOD cDNA clone pSOD1 constitutes the coding region for amino acids 10-153 of hSOD, a single translational stop codon and a 3' untranslated region. Therefore, in the expression vector construct, the base sequence of the region encoding amino acids 1-9 is derived from the published amino acid sequence of hSOD (Jabusch et al., *supra*; Barra et al., *supra*) and synthesized chemically as a part of the variable linker segment (see below).

Construction of Plasmid plot5 - (See Figures 2 and 3)

Plasmid plot1, containing a hybrid trp-lac ("tac") promoter (DeBoer et al., *Proc. Natl. Acad. Sci. USA* - (1983) 80:21-25) was constructed by gel isolating the 180bp HgiA-TaqI fragment of ptrpL1 (Edman et al., *Nature* (1981) 291:503-506) and the 58bp HpaII-EcoRI fragment from pKB268 (Backman and Ptashne, *Cell* - (1978) 13:65-71), and ligating these fragments to pBR322 digested with PstI and EcoRI. The resulting

plasmid was used to transform strain D1210 and clones selected for tetracycline resistance. Plasmid plot3 was constructed by gel isolating the 155bp Fnu4HI-EcoRI fragment of plot1 containing the tac promoter, with the Fnu4HI site being made flush-ended using the Klenow fragment of DNA polymerase I ("pol I K" or "pol. Klen."), and the 18bp EcoRI-PstI polylinker fragment of IIAN7 of the following sequence:

5



15 These fragments were ligated to gel purified pBR322 digested with EcoRI, flush-ended using pol I K, followed by digestion with PstI and gel purified. This ligation mix was used to transform strain D1210, selecting on L-agar plates containing 10µg/ml tetracycline.

Plasmid plot5 was made by first constructing a plasmid containing the IIAN7 polylinker as an EcoRI-PvuII substitution in pBR322. To do this, plasmid IIAN7 was digested with HindIII, made flush-ended by
20 filling in with pol I K and a synthetic, self-complementary, PvuII linker molecule (d(5'-CCAGCTGG-3')) ligated to the above-modified plasmid IIAN7. After digestion with EcoRI and PvuII, the resultant 44bp polylinker fragment (with 4-base overhangs) was gel isolated and cloned into pBR322 as an EcoRI-PvuII substitution.

Plasmid plot3 was digested with EcoRI and after phenol-chloroform extraction and ethanol precipitation,
25 the protruding 5'-ends were made flush-ended by treatment with S1 nuclease (Palmiter, Biochemistry - (1974) 13:3606-3615; Hallewell and Emage, Gene (1980) 9:27-47). After phenol-chloroform extraction and ethanol precipitation, the DNA was digested with ClaI, made flush-ended by pol I K, and the 237bp fragment containing the tac promoter isolated by preparative polyacrylamide gel electrophoresis. This flush-ended tac promoter fragment was then inserted at the PvuII site of the pBR322 polylinker plasmid (see Fig. 3) and
30 clones obtained in which the tac promoter directed transcription towards the β-lactamase gene of pBR322.

Construction of plot5 Derivatives Expressing r-hSOD

The synthetic DNA molecules F(26), C(16), B(31), D(11), E(13) and 4(24) shown in Fig. 1, were
35 synthesized by the phosphoramidite method. The single strand 4(24) was prepared by using all four bases, at each site where X is indicated. Furthermore, silica was withdrawn from the synthesis of the 24mer, such that single-stranded 21mers, 22mers, and 23mers are obtained in addition to the 24mers. After removal from the silica support, the four mixtures are combined in appropriate proportions to provide for equimolar amounts of each of the possible single strands. This mixture was treated as a single product in the
40 subsequent steps.

Molecules F(26), C(16), B(31) and D(11) were mixed together in equimolar amounts and 10µg phosphorylated using T4 polynucleotide kinase. After phenol-ether extraction, the additional non-phosphorylated synthetic DNA molecules 4(24) and E(13) were added, such that all fragments were equimolar. The equimolar mixture contained 13µg of DNA in 133µl of 0.3x kinase buffer.

45 After annealing by cooling at a uniform rate from 70°C to 20°C over 60min, the single strands were ligated together with T4 ligase in 200µl ligation mix at 14°C for 4hrs, phenol-chloroform extracted, ethanol precipitated and the 5'-ends of 4(24) and E(13) phosphorylated using T4 polynucleotide kinase (Maniatis et al., supra). Preparative polyacrylamide gel electrophoresis was used to isolate the completely ligated 53bp material having 5'- and 3'-overhangs.

50 The above purified fragment mixture was then ligated to the 460bp TaqI-PstI segment of the hSOD cDNA as shown in Fig. 1. This segment was itself constructed by isolating the 454bp TaqI-AluI hSOD fragment, making it flush-ended pol I K using and inserting it into plot5 between its EcoRI and SalI sites (see Fig. 3) which had been similarly made flush-ended. After preparation of plasmid DNA from this recombinant, the 460bp TaqI-PstI hSOD fragment was isolated by preparative polyacrylamide gel elec-
55 trophoresis. After extraction and precipitation, the 515bp fragment resulting from the joining of the synthetic fragment to the 460bp TaqI-PstI hSOD fragment was filled in with pol I K (525-528bp) and then digested with SalI and the resulting 519-522bp hSOD fragment isolated by polyacrylamide gel electrophoresis. This fragment was then inserted into plot5 which had been digested with PvuII and SalI and then treated with

alkaline phosphatase. The resulting plasmids were used to transform strain D1210. Recombinants obtained after transformation of strain D1210 were selected on L-agar containing 100µg/ml ampicillin to give a set of clones (designated plot5/SOD) with variable SOD expression.

5 r-hSOD Expression and plot5/SOD Plasmid Selection

For analysis of total *E. coli* proteins by SDS-polyacrylamide gel electrophoresis, overnight cultures were diluted 30-fold into 1ml of L-broth and grown shaking at 37°C for 90min. to an O.D.₅₅₀ of about 0.2. IPTG (isopropylthiogalactoside) was added to a final concentration of 2mM and the cultures incubated an additional 3hrs. After centrifugation, the cell pellet was resuspended in 50µl of gel loading buffer (Laemmli, Nature (1970) 227:680-685) and lysed by repeating the following procedure 3x: Freezing for 1min., boiling for 2min., vortexing for 10sec.

After electrophoresis resolution (Laemmli, *supra*) the protein bands were stained with Coomassie blue and the amount of SOD produced by each clone estimated; these results were then confirmed using Western blots with antibody to authentic human SOD. Over three hundred clones were analyzed and exhibited levels of SOD expression varying from little or none to amounts estimated to be 5-10% of the total soluble cellular protein. Results for six of the over three hundred clones are presented in Table 1, along with the particular sequence for DNA molecule 4(24) as determined by the method of Maxam and Gilbert, *supra*.

Table 1: Sequence and Levels of SOD
Production in *E. coli*

Clone	Sequence: 5'-XXXX ATG GCX ACX	Approximate Weight Percent of Total Protein
pSODx8	AACA A G	5%
pSOD11	GTAT T G	10%
pSODx16	ACAT A A	5%
pSOD5	AATC A A	10%
pSOD16	ATTT T T	10%
pSOD18	AACT A G	7.5%

40 Assays for r-hSOD

For enzyme assays, 100ml cultures were induced as described above. Soluble cell extracts were prepared by vortexing cell pellets 10x20sec. with an equal volume of 0.45-0.5mm diameter glass beads (Braun AG) and an equal volume of assay buffer (50mM Triscacodylate, pH 8.2, 1mM diethylenetriaminepentaacetic acid) in a 1.5ml microfuge tube at 4°C. Cell debris was pelleted by spinning the tube for 1min. in a microfuge (Beckman). The supernatant was dialysed against the assay buffer containing 1mM each of zinc sulfate and copper sulfate for 4hrs at 4°C.

SOD assays were performed using the pyrogallol method (Marklund and Marklund, *supra*). The reaction mixtures employed 0.2mM pyrogallol in assay buffer and reaction rates were determined over a 5min. period using a Hewlett-Packard 8450 spectrophotometer at 420nm. Four different assay samples were prepared: soluble *E. coli* extracts; authentic hSOD; and each of the prior samples pre-incubated with rabbit antibody to authentic hSOD. Each sample was incubated in a cuvette for 1min. at 25°C before adding the pyrogallol and assaying at 25°C. The antibody samples involved a preincubation of 10min. at room temperature in assay buffer with 5µl of antibody. These conditions were found to be sufficient to inactivate 100ng of pure hSOD.

The following Table 2 indicates the results for one of the clones examined (pSODx8):

Table 2: Enzymatic Activity of Human Cu-Zn SOD
Produced in E. Coli (strain D1210 (pSODX8))

<u>Enzyme Preparation</u>	<u>Units SOD/mg Protein</u>
pure Human Cu-Zn SOD	15,384
pSODX8 protein extract	3,017
pSODX8 protein extract preincubated with rabbit anti-human SOD antibody	685
plot5 protein extract	470
plot5 protein extract preincubated with rabbit anti-human SOD antibody	485

These data indicate that approximately 15% of the total soluble cellular protein was hSOD (assuming that the pure human Cu-Zn SOD used as a reference was fully active). Taken together with the electrophoretic data (see above) indicating that 5-10% of total soluble cellular protein migrated as hSOD, it appears that a substantial fraction, probably a majority of the hSOD produced is active.

The correct sequence of the cloned gene was determined by the method of Maxam and Gilbert, supra. In addition, the first twelve amino acids at the N-terminus were determined by automated Edman degradation. The detected sequence of amino acids was as follows:

ALA-THR-LYS-ALA-VAL-(CYS)-VAL-LEU-LYS-GLY-ASP-GLY-

The first ALA residue detected was present at a molar concentration approximately equal to that of the input peptide indicating the absence of a blocked amino terminus. The CYS residue was not detected by the method of amino acid analysis used, but its presence was inferred from the nucleotide sequence.

Thus, the (N-formyl-) methionine was removed from the bacterial expression product and the material had the correct amino acid sequences, i.e., identical to that reported for cytoplasmic hSOD residues 1-10, but the N-terminal ALA residue was not acetylated. Furthermore, the polypeptide made in *E. coli* migrated more slowly than the natural protein in 1% agarose gel (pH 8.6) electrophoresis which detects differences in charge (Corning Universal electrophoresis film, stained according to Clausen, *Immunochemical Technique*, p. 530-531), also indicating lack of acetylation. In addition, analysis of tryptic peptides of both the bacterial hSOD polypeptide and the purified, authentic (acetylated) natural protein revealed that all tryptic peptides were identical, except the bacterial N-terminal peptide which migrated differently, i.e., with a charge expected for a peptide lacking the N-acetyl group.

Expression in Yeast

For transfer of the r-hSOD gene to a yeast vector, the plot5/SOD plasmid clones were screened for an NcoI restriction site at the 5'-end of the coding region. For those plasmids where the variable nucleotides present 5' to the ATG initiation codon are CC, the sequence CCATGG provides an NcoI site. Clones were screened, and one was selected and designated phSOD.

The plasmid phSOD was digested with NcoI and Sall and a 550bp fragment obtained, which included 1

nucleotide untranslated at the 5'-terminus and the entire coding region for hSOD. pPGAP (a yeast expression vector carrying the GAP promoter, prepared as described below) was digested with NcoI and SalI followed by treatment with alkaline phosphatase, and the SalI-NcoI fragment substituted for the NcoI-SalI fragment in pPGAP to provide pPGAPSOD. BamHI digestion of pPGAPSOD resulted in a 2kb fragment
 5 which was gel isolated and inserted into the BamHI site of pC1/1 and pC1/1 GAL4/370, to yield plasmids pC1/1GAPSOD and pC1/1GALGAPSOD, respectively.

Plasmid pC1/1 is a derivative of pJDB219 (Beggs, Nature (1978) 275:104) in which the region corresponding to bacterial plasmid pMB9 in pJDB219 was replaced by pBR322 in pC1/1. For preparing an expression vector having secretory and processing signals, see U.S. application serial no. 522,909. Plasmid
 10 pC1/1GAL4/370, a regulatable yeast expression vector containing the GAL1/GAL10 regulatory region (controlled by the GAL4 gene expression product) is prepared as described below.

Plasmids pC1/1GAPSOD and pC1/1GALGAPSOD were transformed into yeast strain 2150-2-3 (available from Lee Hartwell, University of Washington) as described previously (Hinnen et al. Proc. Natl. Acad. Sci. USA (1978) 75:1929-1933), with the results of expression set forth in the following Table 3.

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Table 3: Expression of Human SOD in Yeast Strain 2150

20	Plasmid	Carbon Source	SOD ² μg/mg protein
25	pC1/1	g, L ¹	0
	pC1/1GAPSOD		148
	pC1/1GALGAPSOD	g, L gal	0.4 68

30

¹ All cultures grown in Minus Leucine media with 2% lactic acid, 3% glycerol with or without 2% galactose to late log or early stationary phase.

35 ² Determined by RIA.

hSOD levels were measured using a standard radioimmunoassay with iodinated authentic hSOD as
 40 standard. Constitutive synthesis from the GAP promoter leads to very high levels of hSOD production, of the order of 10-30% of the total cell protein. The induction with galactose works almost as well, yielding about 7% of the cell protein as hSOD.

When hSOD is produced at these high levels, it is usually necessary to provide zinc and copper ion to the product protein as a prosthetic group in order to recover full enzymatic, i.e., catalytic, activity, e.g., by
 45 dialysis against 1mM solutions of both zinc and copper sulfate. Alternatively, zinc and/or copper ion may be included in ten growth media; this method also provides a means of selecting for strains producing high levels of hSOD and/or avoiding the loss of plasmid vectors expressing hSOD in otherwise non-selective media.

50 Construction of pPGAP

pGAP1, a plasmid prepared by insertion of a HindIII fragment containing the GAPDH gene GAP49 (Holland and Holland, J. Biol. Chem. (1979) 254:5466-5474) inserted in the HindIII site of pBR322, was digested with HinfI and a 500bp fragment isolated. The fragment was resected with Bal31 to remove about
 55 50 or 90bp, followed by ligation with HindIII linkers and digestion with HindIII. pBR322 was digested with HindIII, followed by treatment with alkaline phosphatase and the about 450 or 410bp fragment inserted to provide pGAP128.

pGAP128 was digested with HindIII, the fragment made blunt-ended with the Klenow fragment and

dNTPs and the resulting 450bp fragment isolated by gel electrophoresis. This fragment was inserted into SmaI digested plot5, which had been treated with alkaline phosphatase, to provide plasmid plot5pGAP128, which contained about -400 to +27bp of the GAPDH promoter and coding region.

Yeast expression vector pPGAP having a polyrestriction site linker between the GAPDH terminator and short promoter region was prepared as follows. Plasmid plot5pGAP128 was digested with BamHI and TaqI to yield an approximately 390bp BamHI-TaqI fragment having the -400 to -26bp of the GAPDH promoter. The BamHI-TaqI fragment was ligated to a synthetic fragment containing -25 to -1 bp of the GAPDH promoter and several restriction sites including NcoI and having the following sequence:

10

$$\begin{array}{cccccccccccccccc} \text{CGA}_2\text{TA}_3 & (\text{CA})_3 & \text{TA}_3\text{CA}_3 & \text{CACCATG}_3 & \text{A}_2\text{T}_2 & \text{CGT}_2\text{AG}_2 \\ \text{T}_2\text{AT}_3 & (\text{GT})_3 & \text{AT}_3\text{GT}_3 & \text{GTGGTAC}_3 & \text{T}_2\text{A}_2 & \text{GCA}_2\text{TC}_2 & \text{AGCT} \end{array}$$

to provide a BamHI-Sall fragment, which was digested with BamHI and Sall and used to replace the BamHI-Sall fragment of BamHI-Sall digested pBR322 treated with alkaline phosphatase. After ligation, the plasmid pGAPNRS was obtained which was digested with BamHI and Sall to provide a 400bp BamHI-Sall fragment which was gel isolated. This fragment was ligated to an about 900bp Sall-BamHI fragment containing the GAPDH terminator region and a short segment of 3' coding region and the resulting 1.4kb BamHI-BamHI fragment digested with BamHI. The Sall-BamHI GAPDH terminator fragment was obtained by Sall and BamHI digestion of pGAP2, a plasmid prepared by insertion of an about 3.3kb BamHI fragment containing the GAPDH gene GAP49 (Holland and Holland, *supra*) into the BamHI site of pBR322. Plasmids pGAP2 and pGAP1 were obtained as follows: A yeast gene library was prepared by inserting fragments obtained after partial digestion of total yeast DNA with restriction endonuclease Sau3A in lambda-phage Charon 28 (Blattner et al., *Science* (1977) 196:161-169). The phage library was screened with DNA complementary to the yeast GAPDH mRNA and the yeast GAPDH gene from one of these clones was subcloned as either an about 3.3kb BamHI fragment in the BamHI site of pBR322 (pGAP-2) or as an about 2.1kb HindIII fragment in the HindIII site of pBR322 (pGAP-1).

pBR322 was digested with EcoRI and Sall, the termini blunt-ended and ligated to BamHI linkers, followed by BamHI digestion and the BamHI-BamHI 3.8kb fragment gel isolated, recircularized by self-ligation, cloned and designated pBRΔR1-Sal. The 1.4kb BamHI-BamHI fragment was inserted into the BamHI-digested, alkaline phosphatase treated pBRΔR1-Sal vector to provide the plasmid pPGAP of about 5.3kb with the orientation in the opposite direction of the amp^r.

35 Construction of GAL regulated containing plasmids.

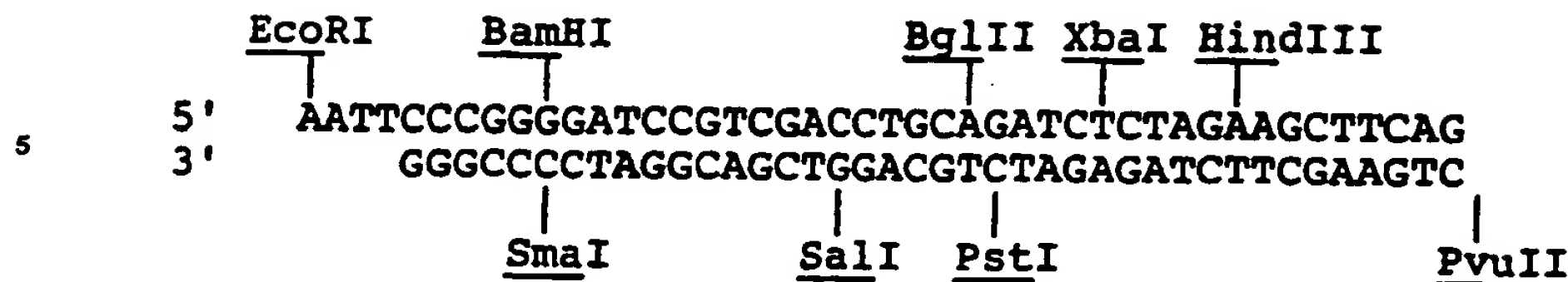
Plasmid pLGSD5 is prepared as described in Guarente et al., (1982) *supra*. The plasmid was manipulated as follows: After restriction with XhoI, the overhangs were filled in with the Klenow fragment of DNA polymerase I ("Klenow fragment"), ligated with EcoRI linkers (GGAATTCC) and then completely digested with EcoRI and Sau3A to provide a 370bp fragment which was isolated by gel electrophoresis and included the intergenic sequence between GAL1 and GAL10 genes of yeast, and provides for the GAL4 regulation sequence of the GAL1 and GAL10 genes.

This fragment was inserted into pBR322 which had been completely digested with EcoRI and BamHI, followed by treatment with alkaline phosphatase to prevent oligomerization resulting in plasmid pBRGAL4.

Plasmid pBRGAL4 was completely digested with Sau3A, the overhangs filled in with the Klenow fragment, and the resulting blunt-ended fragment ligated with Sall linkers (CGTCGACG), followed by digestion with Sall and XhoI. The resulting 370bp fragment was isolated by gel electrophoresis. This fragment has the original 370bp yeast GAL4 regulator sequence with XhoI and Sall termini.

The fragment was then cloned in the plasmid plot5. plot5 was prepared by inserting the 40bp polylinker fragment of the following sequence

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into pBR322 as an EcoRI-PvuII substitution followed by insertion of the trp-lac promoter (Russell and Bennett, Gene (1982) 20:231-245) into the PvuII site with transcription oriented toward the polylinker sequence. plot5 was completely digested with SalI, followed by treatment with alkaline phosphatase and the 370bp fragment inserted into plot5 to provide plasmid plot5GAL4/370. This plasmid was then completely digested with BamHI and SalI to reproduce the individual fragment extended by 8bp of the polylinker fragment. This fragment was then ligated into pC1/1, which had been completely digested with BamHI and SalI followed by treatment with alkaline phosphatase to prevent recircularization. The resulting plasmid was designated pC1/1GAL4/370. The BamHI-SalI fragment is located in the pBR322 portion of the vector pC1/1.

The phSOD polypeptide made in yeast was shown to be identical to the natural human protein. Migration of hSOD made in yeast was identical to the protein in both polyacrylamide gel electrophoresis (with and without sodium dodecyl sulfate) and in agarose gel electrophoresis (see above). Moreover, when highly purified yeast polypeptide was subjected to twelve cycles of Edman degradation, the sequence was the same as that reported for the human protein (residues 1-10) made in E. coli set forth above. The level of detection, however, was only 5 to 10% of the expected level on a molar basis relative to protein input. This reduced detection level indicated that the N-terminal amino acid was blocked, i.e., probably acetylated. The result was confirmed by comparative analysis of tryptic peptides derived from yeast-produced hSOD and authentic acetylated human material which showed that all the expected tryptic proteins were identical in the two samples including the N-terminal one, thus indicating the presence of acetylated N-terminal ALA in the yeast expression product.

Isolation of the Human SOD Gene

To isolate the human SOD gene, a bacteriophage lambda library representative of the human genome (R. Lawn et al. (1978) Cell 15:1157-1174) was screened with a radiolabelled DNA probe made from the human SOD cDNA. One million phage plaques were screened, and 13 positively hybridizing plaques were purified. Restriction endonuclease analysis of the phage DNAs indicated that there are at least 5 different genes, suggesting that there are other SOD related genes and gene products. One candidate for such a gene is the recently discovered extracellular Cu/Zn SOD (S. Marklund, (1982) Proc. Natl. Acad. Sci. USA 79:7634-7638). To distinguish the authentic cytoplasmic Cu/Zn SOD gene from the related ones we used synthetic DNA probes 5'-AATGCTTCCCCACACC-3' and 5'-CTCAGTTAAATGTCTGTTTG-3' corresponding to amino acid residues 19-26 and nucleotides 193-213 3' from the terminator codon in the 3' untranslated region, respectively. Only one of the 13 genomic DNAs hybridized with these probes, indicating that it is the authentic human cytoplasmic SOD gene. This was confirmed by DNA sequence analysis of the N-terminal region, as shown in Fig. 5 where the amino acid sequence determined by protein sequencing is confirmed. This also shows that no preprotein exists for SOD since an in-frame termination codon exists nine nucleotides 5' from the initiator methionine codon. As shown in Figure 5, the human Cu/Zn SOD gene contains intervening sequences. The map of the SOD gene shown in Figure 6 indicates that there is more than one intervening sequence.

Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

Claims

1. A method for producing an acetylated human superoxide dismutase, said method comprising introducing into yeast a DNA sequence encoding said human superoxide dismutase and having an acetylation signal sequence at its 5'-end encoding for glycine or alanine at the first position followed by a polar amino acid at the second position, which acetylation signal sequence causes the yeast host to acetylate

the N-terminal amino acid on the polypeptide.

2. A method as in claim 1, wherein the polar amino acid is selected from the group consisting of aspartate, serine and threonine.

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Revendications

1. Un procédé pour produire une superoxyde dismutase humaine acétylée, cette méthode comportant l'introduction dans une levure d'une séquence d'ADN codant pour cette superoxyde dismutase humaine et comportant à son extrémité 5' une séquence-signal d'acétylation codant pour la glycine ou l'alanine, en première position, suivie par un acide aminé polaire, en seconde position, ladite séquence-signal d'acétylation causant la levure hôte à acétyler l'acide aminé N-terminal sur le polypeptide.
2. Un procédé selon la revendication 1, dans lequel l'acide aminé polaire est choisi parmi le groupe consistant en aspartate, sérine et thréonine.

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Patentansprüche

1. Verfahren zur Herstellung einer acetylierten Human-Superoxiddismutase, das das Einführen einer die Human-Superoxiddismutase kodierenden und an ihrem 5'-Ende eine ein Glycin oder Alanin in der ersten Position, gefolgt von einer polaren Aminosäure in der zweiten Position, kodierende Acetylierungs-Signalsequenz aufweisenden DNA-Sequenz in Hefe einschließt, wobei die Acetylierungs-Signalsequenz den Hefewirt zum Acetylieren der N-terminalen Aminosäure des Polypeptids veranlaßt.
2. Verfahren nach Anspruch 1, in dem die polare Aminosäure aus der von Aspartat, Serin und Threonin gebildeten Gruppe ausgewählt ist.

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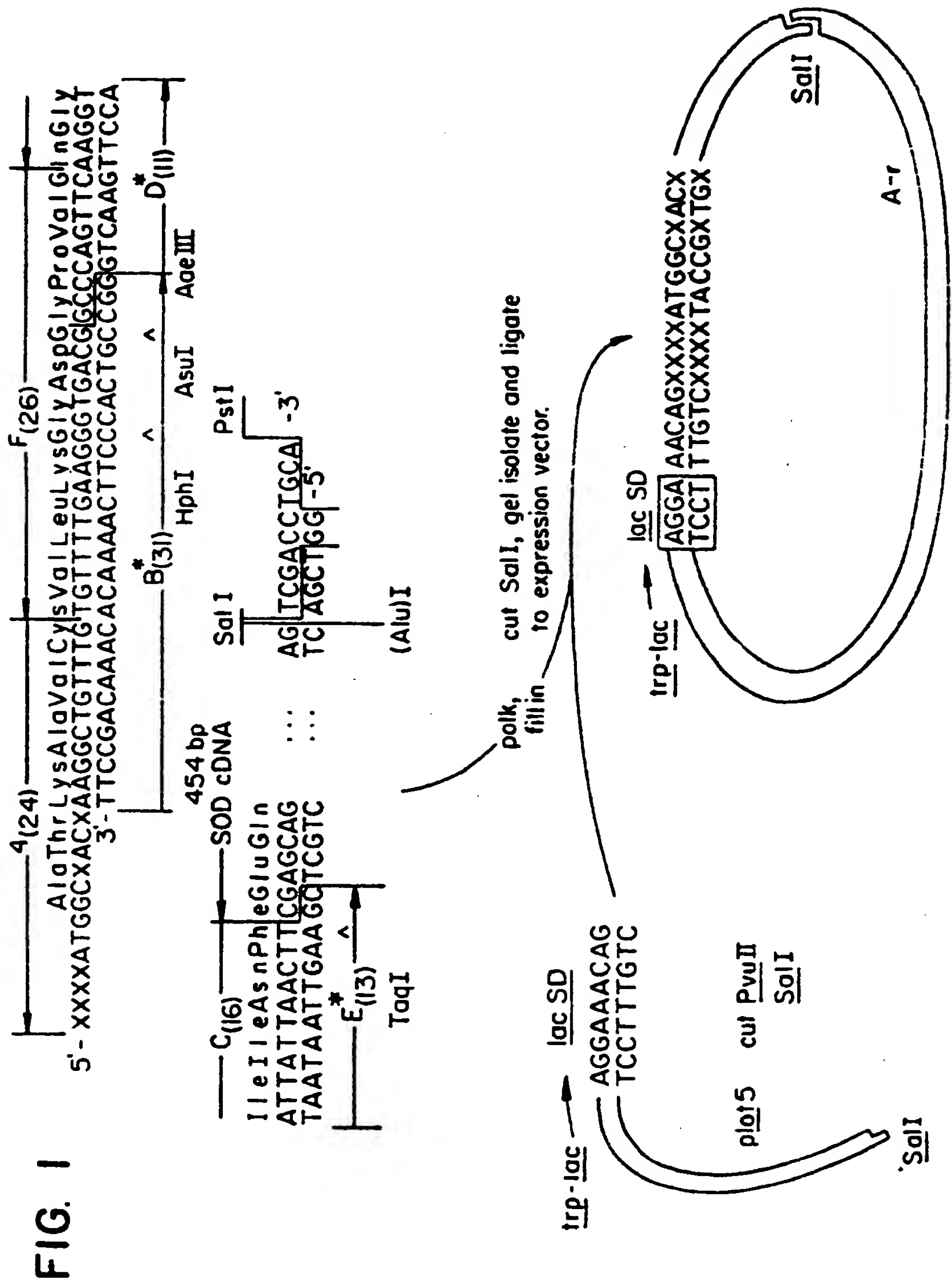
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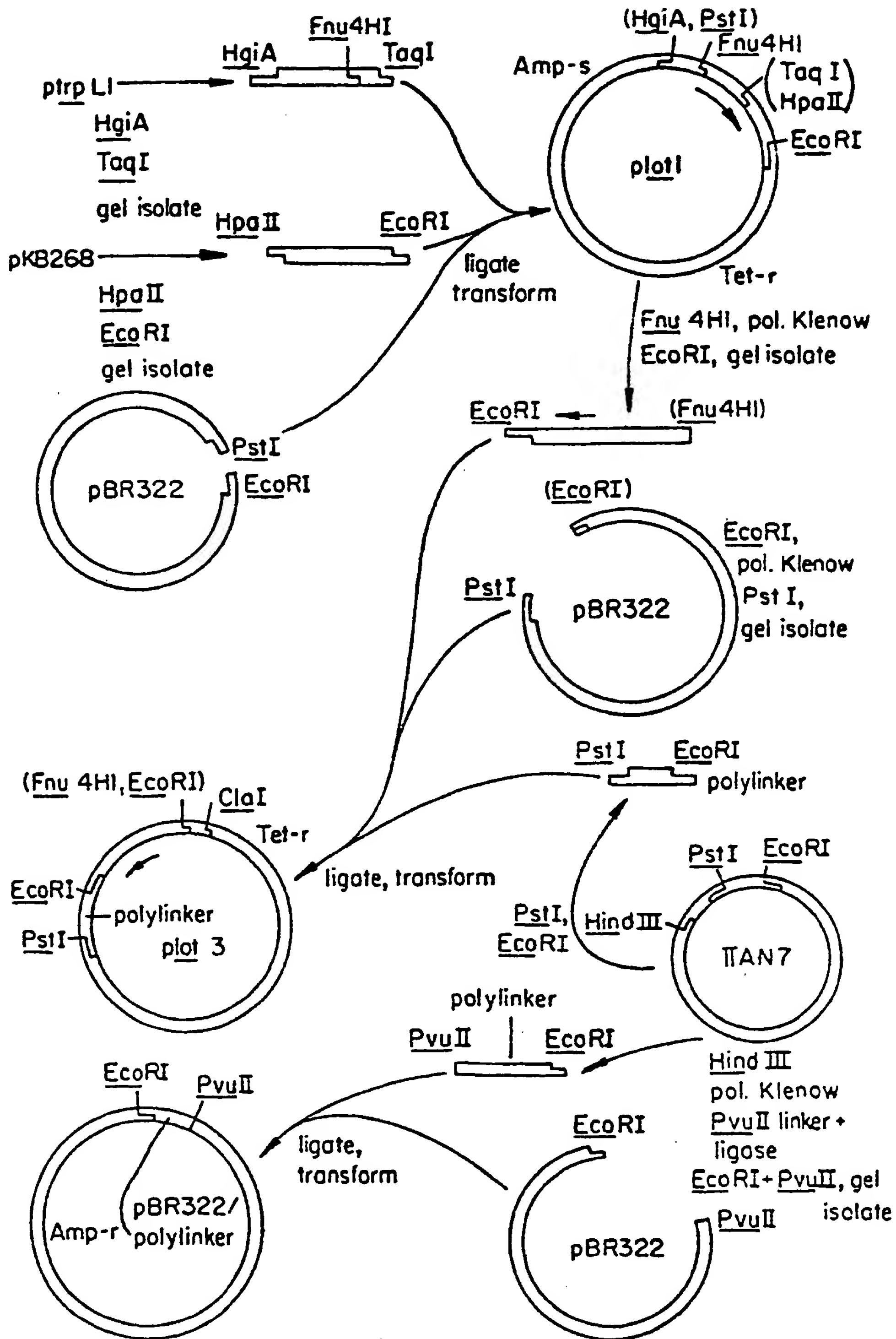


FIG. 2

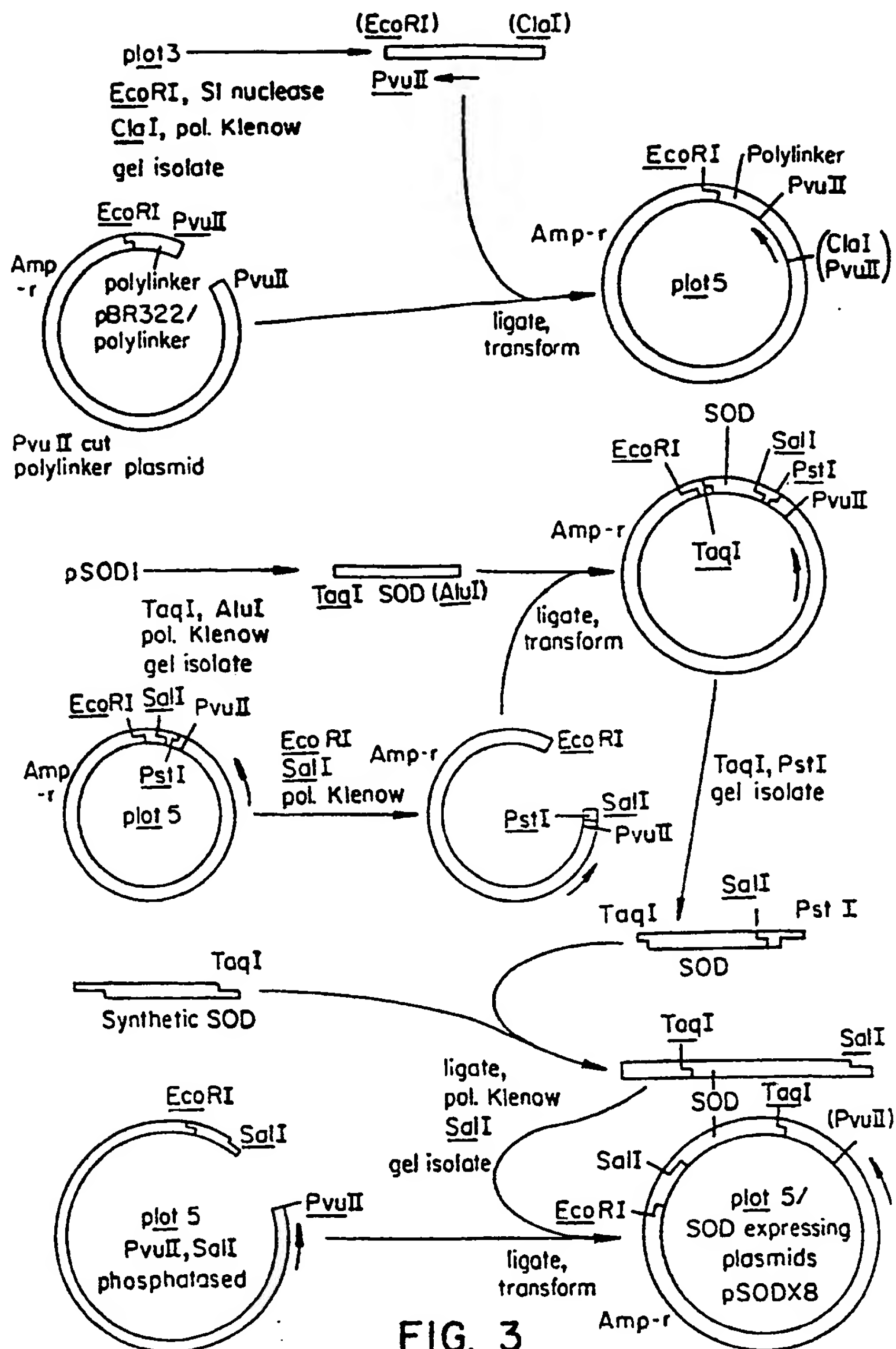


FIG. 3

Translation of hsod

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10                                20
gly asp gly pro val gln gly ile ile asn phe glu gln lys glu
GGC GAC GGC CCA GTG CAG GGC ATC ATC AAT TTC GAG CAG AAG GAA
      *Sau 96 I                                *TaqI

30
ser asn gly pro val lys val trp gly ser ile lys gly leu thr
AGT AAT GGA CCA GTG AAG GTG TGG GGA AGC ATT AAA GGA CTG ACT
      *Sau 96 I

40                                50
glu gly leu his gly phe his val his glu phe gly asp asn thr
GAA GGC CTG CAT GGA TTC CAT GTT CAT GAG TTT GGA GAT AAT ACA

60
ala gly cys thr ser ala gly pro his phe asn pro leu ser arg
GCA GGC TGT ACC AGT GCA GGT CCT CAC TTT AAT CCT CTA TCC AGA

70                                80
lys his gly gly pro lys asp glu glu arg his val gly asp leu
AAA CAC GGT GGG CCA AAG GAT GAA GAG AGG CAT GTT GGA GAC TTG

90
gly asn val thr ala asp lys asp gly val ala asp val ser ile
GGC AAT GTG ACT GCT GAC AAA GAT GGT GTG GCC GAT GTG TCT ATT

100                                110
glu asp ser val ile ser leu ser gly asp his cys ile ile gly
GAA GAT TCT GTG ATC TCA CTC TCA GGA GAC CAT TGC ATC ATT GGC

120
arg thr leu val val his glu lys ala asp asp leu gly lys gly
CGC ACA CTG GTG GTC CAT GAA AAA GCA GAT GAC TTG GGC AAA GGT

130                                140
gly asn glu glu ser thr lys thr gly asn ala gly ser arg leu
GGA AAT GAA GAA AGT ACA AAG ACA GGA AAC GCT GGC AGT CGT TTG

150                                153
ala cys gly val ile gly ile ala gln OC
GCT TGT GGT GTA ATT GGG ATC GCC CAA TAA ACATTCCTTGCTGTAGTCT

GAGGCCCTTAACATCATCTGTTATCCTGCTAGCTC-
      *AluI

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Translated Mol. Weight = 14893.03

FIG. 4

CTGCAGCGTCTGGGGTTTCCGTTGCAGTCCTCGGACCAACGACCTCGGCGTGCCCTAGCGAGTT Met ATG
Ala Thr Lys Ala Val Cys Val Leu Lys Gly Asp Gly Pro Val Gln Gly Ile Ile
GCG ACG AAG GCC GTG TGC GTG CTG AAG GGC GAG GGC CCA GTG CAG GGC ATC ATC
Asn Phe Glu Gln Lys GCAAGGGCTGGGACGGAGGCTTG
AAT TTC GAG CAG AAG
EXON<----->INTRON

FIG. 5

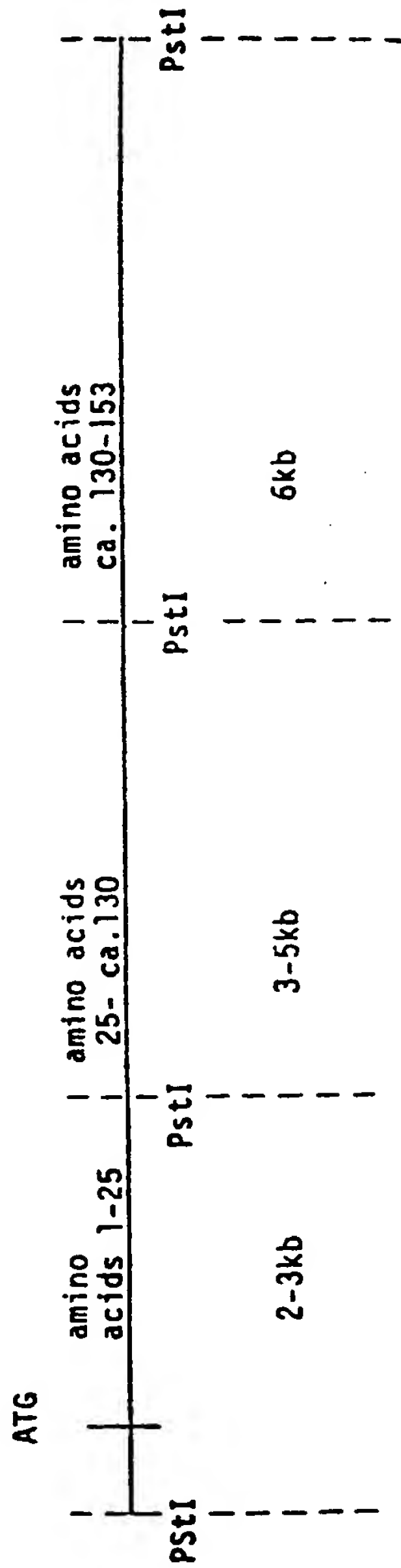


FIG. 6